

## ORIGINAL PAPER

**AUGMENTED IMMUNOEXPRESSION OF SURVIVIN CORRELATES WITH PARAMETERS OF AGGRESSIVENESS IN PROSTATE CANCER**MARIAN DANILEWICZ<sup>1</sup>, OLGA STASIKOWSKA-KANICKA<sup>2</sup>, MAŁGORZATA WĄGROWSKA-DANILEWICZ<sup>2</sup><sup>1</sup>Department of Pathology, Medical University of Lodz, Lodz, Poland<sup>2</sup>Department of Nephropathology, Medical University of Lodz, Lodz, Poland

The clinical significance of the immunoeexpression of survivin in prostate cancer and its correlation with the biological aggressiveness of prostate cancer remains unclear. Therefore, the present study was undertaken to compare the immunoeexpression of survivin in prostate cancer (PCa) and benign prostatic hyperplasia (BPH) as well as to determine whether this immunoeexpression could correlate with Gleason score, proliferation activity and prostate specific antigen (PSA) levels. The prostate needle biopsies from 28 patients with elevated serum PSA levels were studied. As a control, 12 needle biopsies of prostate diagnosed as BPH were used. The immunoeexpression of survivin was evaluated semiquantitatively, whereas the Ki-67 index was assessed quantitatively. The immunoeexpression of survivin and Ki-67 in epithelial cells in the prostate cancer group was significantly increased as compared to BPH cases. In the prostate cancer group there were positive significant correlations between the immunoeexpression of survivin and Gleason score as well as Ki-67 antigen. The correlation between the immunoeexpression of survivin and PSA levels was also positive, but it did not reach statistical significance. In conclusion, we can confirm that in prostate cancer the immunoeexpression of survivin is augmented as compared to BPH and positively correlated with parameters of tumor aggressiveness.

**Key words:** prostate cancer, survivin, Ki-67, PSA.

**Introduction**

Worldwide, prostate cancer (PCa) is the second most frequently diagnosed cancer in males [1-4]. There are over 540 000 new cases diagnosed each year worldwide [5]. It occupies second place in mortality in Poland and first place in the USA [6, 7].

The classic, widely used prognostic factors for prostate cancer are prostate specific antigen (PSA) serum levels and histological grading – Gleason score and staging represented by the TNM system. Prostate specific antigen serum level is widely used for PCa screening and has resulted in an overall decrease in PCa metastasis and death. However, this screening has also led to over-detection and overtreatment as

elevations in PSA levels can also be present as a result of infection, chronic inflammation or benign prostatic hyperplasia (BPH) [1]. Thus, searching for new methods of therapy and prognosis becomes a key task for many researchers around the world. Recently, the prognostic role of angiogenesis was investigated [8].

Prostate cancer is slow-growing malignancy that is characterized by an imbalance in the rates of cell division and cell death which mainly depends on apoptosis [1, 9]. Survivin is a new and the smallest member of the inhibitor of apoptosis family and is expressed predominantly in fetal tissue but is also found in many human malignancies. It is a 142-amino acid, 16.5-kDa protein coded by a single-copy gene on the human 17q25 chromosome [10-12]. Although the

immunoexpression of survivin in prostate cancer has been reported, the clinical significance of its expression and correlation with the biological aggressiveness of prostate cancer remains unclear [13, 14]. Especially the correlation of expression of survivin and Gleason score is controversial [1, 13, 15, 16].

Therefore, the present study was undertaken to compare the immunoexpression of survivin in PCa and BPH as well as to determine whether this immunoexpression could correlate with Gleason score, proliferation activity and PSA levels.

## Material and methods

### Patients

The investigation was confined to study prostate needle biopsies from 28 patients diagnosed in the Department of Pathology, Medical University of Lodz (years 2007-2012). In all cases indication for the biopsy was elevated serum PSA level. Mean age of the patients was  $68.85 \pm 5.91$ , and PSA level varied from 4.02 ng/ml to 103 ng/ml, mean  $34.62 \pm 35.2$ . All patients of this group had prostate cancer diagnosed using light microscopy and standard hematoxylin-eosin stain. As a control, 12 needle biopsies of prostate diagnosed as BPH were used. In all these patients the serum level of PSA was within normal limits.

### Immunohistochemistry

Paraffin-embedded tissue sections were mounted onto SuperFrost slides, deparaffinized, then treated in a microwave oven in a solution of TRS (Target Retrieval Solution, pH 6.0, Dako) for 30 minutes ( $2 \times 6$  minutes 360 W,  $2 \times 5$  minutes 180 W,  $2 \times 4$  minutes 90 W) and transferred to distilled water. Endogenous peroxidase activity was blocked by 0.3% hydrogen peroxide in distilled water for 30 minutes,

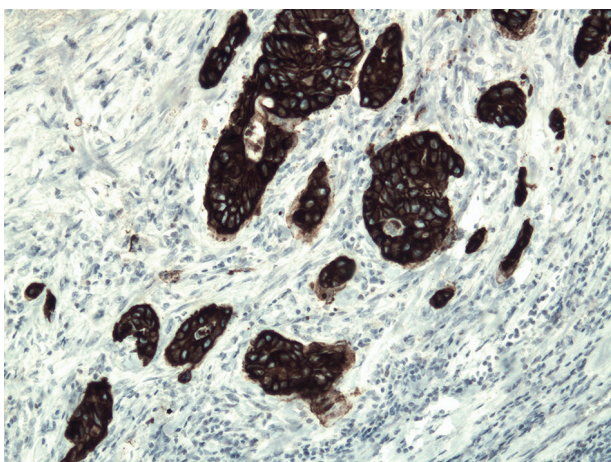


Fig. 1. Positive control for survivin (adenocarcinoma of the colon). Magnification 100×

and then sections were rinsed with Tris-buffered saline (TBS, Dako, Denmark) and incubated all night with monoclonal anti-human survivin antibody (Abcam; dilution 1 : 300) and 30 minutes with mouse monoclonal anti-human Ki 67 antibody (Dako; dilution 1 : 100). Immunoreactive proteins were visualized using the EnVision-HRP kit (Dako, Carpinteria, CA, USA) according to the instructions of the manufacturer. Visualization was performed by incubating the sections in a solution of 3,3'-diaminobenzidine (DakoCytomation, Denmark). After washing, the sections were counter-stained with hematoxylin and coverslipped. For each antibody and for each sample a positive and negative control were processed. As a positive control, according to the instructions of the manufacturer, adenocarcinoma of the colon was used (Fig. 1). Negative controls were carried out by incubation in the absence of the primary antibody and always yielded negative results (Fig. 2).

In each specimen staining intensity of survivin in epithelial cells was recorded semiquantitatively by two independent observers in 7-10 adjacent high power fields and graded 0 (staining not detectable), 1 (weak immunostaining), 2 (moderate immunostaining intensity) or 3 (strong staining). The mean grade was calculated by averaging grades assigned by the two authors and approximating the arithmetic mean to the nearest unity.

### Morphometry

Ki-67 positive cells were evaluated using a computer image analysis system consisting of a PC computer equipped with a Pentagram graphic tablet, Indeo Fast card (frame grabber, true-color, real-time), produced by Indeo (Taiwan), and color TV camera Panasonic (Japan) coupled with a Carl Zeiss microscope (Germany). This system was programmed (MultiScan 8.08 software, produced by Computer

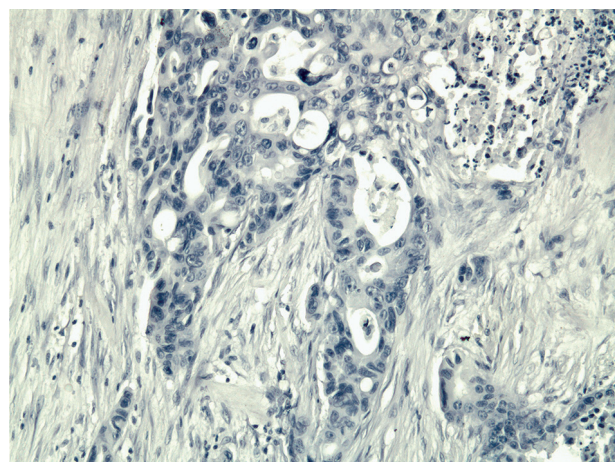


Fig. 2. Negative control for survivin (adenocarcinoma of the colon). Magnification 100×

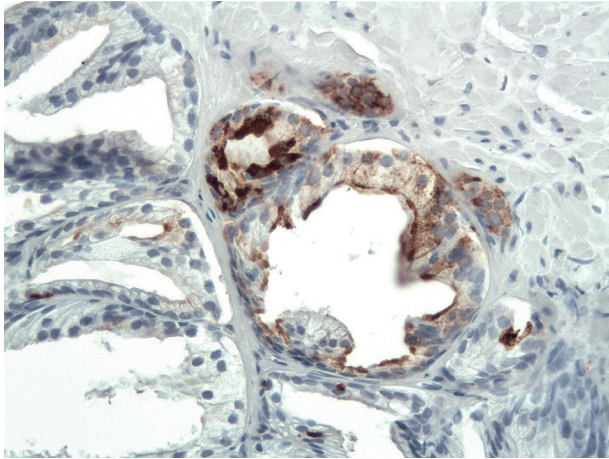


Fig. 3. Benign prostatic hyperplasia. Weak immunoreactivity of survivin in the cytoplasm of epithelial cells. Magnification 200×

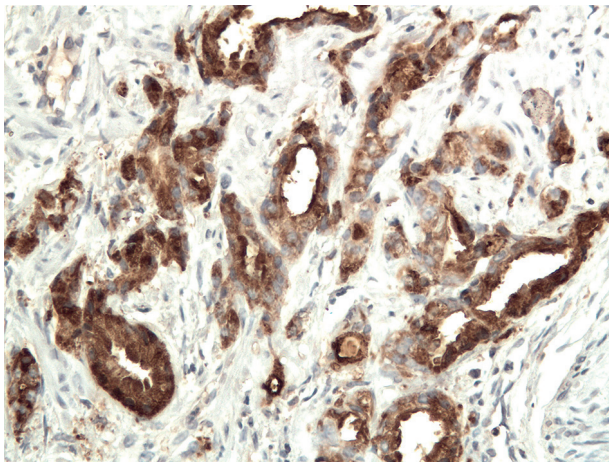


Fig. 4. Prostate cancer. Strong immunoreactivity of survivin in the cytoplasm of epithelial cells. Magnification 200×

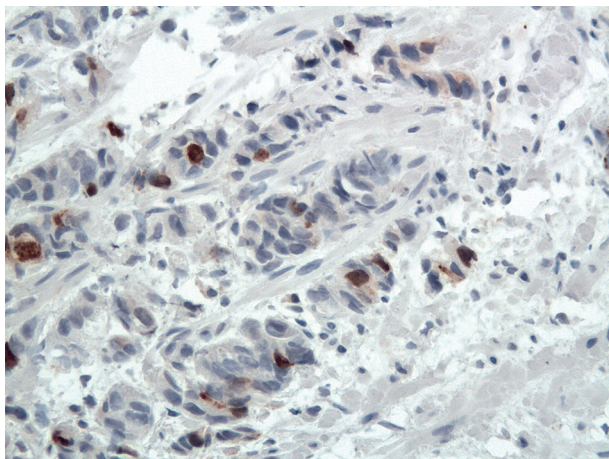


Fig. 5. Prostate cancer. Nuclear immunoreactivity of survivin. Magnification 200×

Scanning Systems, Poland) to calculate the number of objects (semiautomatic function).

The percentage of Ki-67 positive cells was estimated by counting 100 cells in five monitor fields

(0.029 mm<sup>2</sup> each), marking immunopositive cells (semiautomatic function), so that in each case 500 cells were analyzed.

### Statistical methods

All values were expressed as a mean ± SD (standard deviation). The differences between groups were tested using the Mann-Whitney U test. Correlation coefficients were calculated using Spearman's method. Results were considered statistically significant if  $p < 0.05$ .

### Results

In BPH biopsies we found 8 of 12 cases with a positive survivin reaction.

The immunoreactivity of survivin protein was also positive in 26 of 28 cases of prostatic cancer. Four of these cases were low risk PCa (PSA level < 10 ng/ml and Gleason score < 6); 2 of them were survivin negative. Twenty-four cases were survivin-positive high-risk PCa. In BPH the immunoreactivity of survivin was exclusively membranous/cytoplasmic (Fig. 3). Similarly, in most cases of prostate cancer the immunoreactivity of survivin was membranous/cytoplasmic (Fig. 4), although in some patients a nuclear reaction for survivin was seen (Fig. 5). In both BPH and prostate cancer groups the immunoreactivity of Ki-67 was exclusively nuclear (Figs. 6, 7).

The semiquantitative and morphometric data of the immunoreactivity of survivin and Ki-67 antigen appear in Table I. The immunoreactivity of survivin and Ki-67 in epithelial cells in the prostate cancer group was significantly increased as compared to BPH cases.

The correlations between immunoreactivity of survivin and Gleason score, PSA levels and Ki-67 antigen are given in Table II. In the prostate cancer group there were positive significant correlations between immunoreactivity of survivin and Gleason score as well as Ki-67 antigen. The correlation between immunoreactivity of survivin and PSA levels was also positive, but it did not reach statistical significance. In BPH patients all these correlations were weak and not significant.

### Discussion

Survivin, an anti-apoptotic protein, inhibits the processing of initiator caspase-9 as well as caspase-3 and caspase-7, terminal effectors of apoptosis. Additionally, survivin counteracts pro-apoptotic stimuli induced by interleukin-3, Fas, Bax, tumor necrosis factor and anticancer drugs [13, 17-21]. Besides the anti-apoptotic properties, survivin regulates cell division [22-24]. In prostate cancer aberrantly activated signal transducer and activator of transcription 3

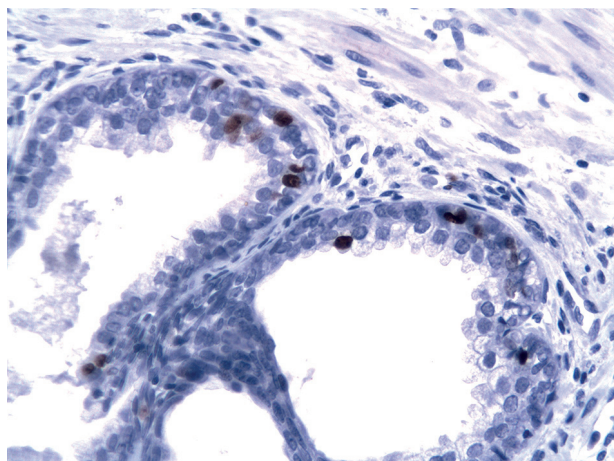


Fig. 6. Weak immunopositivity of Ki-67 protein in benign prostatic hyperplasia. Magnification 200 $\times$

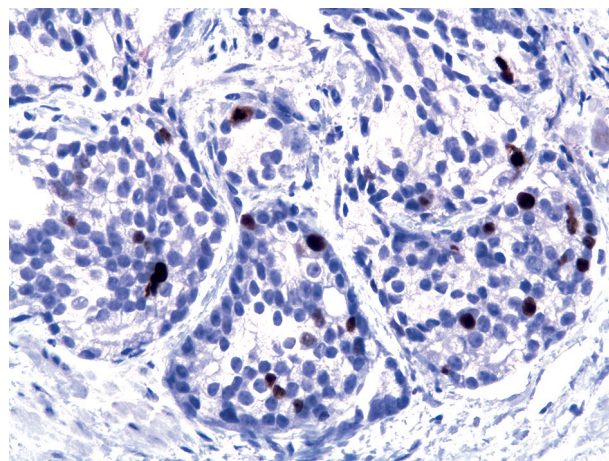


Fig. 7. Numerous nuclei marked as Ki-67 positive in patient with prostate cancer. Magnification 200 $\times$

(Stat3) is probably involved in direct activation of survivin and other proto-oncogenes [25].

In the present study we confirmed the findings of Shariat *et al.* [26] that the immunopositivity of survivin is significantly increased in prostate cancer patients as compared to controls, whereas Kishi *et al.* revealed that survivin mRNA was in PCa significantly higher than that found in control tissues [13]. Koike *et al.* quantified the gene expression levels of survivin and its isoforms in PCa and BPH and found that in prostate biopsy samples, the survivin expression in PCa was significantly greater than that in BPH or PCa after androgen deprivation therapy [15]. In the results of other investigators, plasma-derived exosomal survivin was also significantly increased in the PCa-derived samples, compared to those from BPH [1]. Similarly, in the paper of Bialkowska-Hobrzanska *et al.* survivin mRNA was in PCa up to 13.5-fold higher as compared to control tissue [16].

As might be expected, in our study the percentage of Ki-67 positive nuclei was significantly higher in prostate cancer patients in comparison with BPH individuals. However, the most important findings in the present paper are correlative data. We revealed a strong positive correlation between the immunopositivity of survivin and Gleason score in the PCa group. This result supports the observations that survivin correlates with the parameters of aggressiveness in prostate cancer. In the study of Kishi *et al.* in PCa

Table I. Semiquantitative data of immunopositivity of survivin and morphometric data of Ki-67 score in prostate cancer and controls

NUMBER OF CASES	CONTROLS (BPH) (N = 12)	PROSTATE CANCER (N = 28)	P VALUE
Survivin (mean score)	0.92 $\pm$ 0.82	2.25 $\pm$ 1.76	< 0.02
Ki-67 (%)	0.97 $\pm$ 0.53	12.32 $\pm$ 5.25	< 0.001

samples survivin expression was significantly higher in cancers with a high Gleason score, high pathological T stage, positive surgical margin and vessel invasion [13]. Similarly, Koike *et al.* and Shariat *et al.* found that survivin expression was also associated with high Gleason score [15, 26]. Contrary to us and cited authors, Khan *et al.* [1] and Bialkowska-Hobrzanska *et al.* [16] did not reveal any correlations between survivin and Gleason score. These differences may depend on methodological reasons, as to evaluate survivin these researchers used various methods.

Moreover, we found a significant positive correlation between immunopositivity of survivin and the Ki-67 index. Similarly, other authors demonstrated that survivin expression is positively related to PCa proliferation activity measured by the PCNA labeling index [13]. However, to our best knowledge in

Table II. Correlations between immunopositivity of survivin and Gleason score, PSA levels and Ki-67 score in prostate cancer as well as in controls

CORRELATION BETWEEN	CONTROLS (BPH) (N = 12)	PROSTATE CANCER (N = 28)
Immunopositivity of survivin and Gleason score	–	r = 0.61, p < 0.005
Immunopositivity of survivin and PSA levels	r = 0.49, p = 0.1 (NS)	r = 0.36, p = 0.06 (NS)
Immunopositivity of survivin and Ki-67 (%)	r = 0.32, p = 0.31 (NS)	r = 0.54, p < 0.004

NS – not significant

the available literature no data have documented such a relationship using Ki-67 antibody. Finally, we noticed in PCa cases a positive correlation between the immunoexpression of survivin and PSA serum levels, but this relationship was not significant. Interestingly, in a previous study [13] the survivin expression in PCa patients with a short preoperative prostate specific antigen doubling time (less than 2 years) was also significantly higher than those with a moderate (2-4 years) or long (greater than 4 years) doubling time.

In conclusion, we can confirm that in prostate cancer the immunoexpression of survivin is augmented as compared to BPH and positively correlated with parameters of tumor aggressiveness.

*The authors declare no conflict of interest.*

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## References

1. Khan S, Jutzy JM, Valenzuela MM, et al. Plasma-derived exosomal survivin, a plausible biomarker for early detection of prostate cancer. *PLoS One* 2012; 7: e46737.
2. Jemal A, Siegel R, Xu J, et al. Cancer statistics, 2010. *CA Cancer J Clin* 2010; 60: 277-300.
3. Jemal A, Bray F, Center MM, et al. Global cancer statistics. *CA Cancer J Clin* 2011; 61: 69-90.
4. Globa T, Saptefręi L, Ceașu R, et al. Mast cell phenotype in benign and malignant tumors of the prostate. *Pol J Pathol* 2014; 65: 147-153.
5. Schröder FH. Screening for prostate cancer. *Urol Clin North Am* 2003; 30: 239-251.
6. Stawerski P, Wągrowska-Danilewicz M, Stasikowska-Kanicka O, et al. Augmented mast cell infiltration and microvessel density in prostate cancer. *Contemp Oncol (Pozn)* 2013; 17: 378-382.
7. Kaczmarczyk K, Dyduch G, Białas M, et al. Frequency of ERG-positive prostate carcinoma in Polands. *Pol J Pathol* 2013; 64: 175-179.
8. Łuczyńska E, Gasińska A, Wilk W. Microvessel density and expression of VEGF in clinically localized prostate cancer. *Pol J Pathol* 2013; 64: 33-38.
9. Lu S, Liu M, Epner DE, et al. Androgen regulation of the cyclin-dependent kinase inhibitor p21 gene through an androgen response element in the proximal promoter. *Mol Endocrinol* 1999; 13: 376-384.
10. Zhang M, Coen JJ, Suzuki Y, et al. Survivin is a potential mediator of prostate cancer metastasis. *Int J Radiat Oncol Biol Phys* 2010; 78: 1095-103.
11. Altieri DC. Validating survivin as a cancer therapeutic target. *Nat Rev Cancer* 2003; 3: 46-54.
12. Li F. Survivin study: What is the next wave? *J Cell Physiol* 2003; 197: 8-29.
13. Kishi H, Igawa M, Kikuno N, et al. Expression of the survivin gene in prostate cancer: correlation with clinicopathological characteristics, proliferative activity and apoptosis. *J Urol* 2004; 171: 1855-1860.
14. Ambrosini G, Adida C, Altieri DC. A novel anti-apoptosis gene, survivin, expressed in cancer and lymphoma. *Nat Med* 1997; 3: 917-921.
15. Koike H, Sekine Y, Kamiya M, et al. Gene expression of survivin and its spliced isoforms associated with proliferation and aggressive phenotypes of prostate cancer. *Urology* 2008; 72: 1229-1233.
16. Bialkowska-Hobrzanska H, Driman DK, Fletcher R, et al. Expression of human telomerase reverse transcriptase, Survivin, DD3 and PCGEM1 messenger RNA in archival prostate carcinoma tissue. *Can J Urol* 2006; 13: 2967-2974.
17. Tamm I, Wang Y, Sausville E, et al. IAP-family protein survivin inhibits caspase activity and apoptosis induced by Fas (CD95), Bax, caspases, and anticancer drugs. *Cancer Res* 1998; 58: 5315-5320.
18. Swana HS, Grossman D, Anthony JN, et al. Tumor content of the antiapoptosis molecule survivin and recurrence of bladder cancer. *N Engl J Med* 1999; 341: 452-453.
19. Altieri DC. The molecular basis and potential role of survivin in cancer diagnosis and therapy. *Trends Mol Med* 2001; 7: 542-547.
20. Salvesen GS, Duckett CS. IAP proteins: blocking the road to death's door. *Nat Rev Mol Cell Biol* 2002; 3: 401-410.
21. Schimmer AD. Inhibitor of apoptosis proteins: translating basic knowledge into clinical practice. *Cancer Res* 2004; 64: 7183-7190.
22. Chen J, Cui X, Zhou H, et al. Functional promoter -31G/C variant of Survivin gene predict prostate cancer susceptibility among Chinese: a case control study. *BMC Cancer* 2013; 13: 356.
23. Li F, Ambrosini G, Chu EY, Plescia J, et al. Control of apoptosis and mitotic spindle checkpoint by survivin. *Nature* 1998; 396: 580-584.
24. Li F, Altieri DC. The cancer antiapoptosis mouse survivin gene: characterization of locus and transcriptional requirements of basal and cell cycle-dependent expression. *Cancer Res* 1999; 59: 3143-3151.
25. Qin HR1, Kim HJ, Kim JY, et al. Activation of signal transducer and activator of transcription 3 through a phosphomimetic serine 727 promotes prostate tumorigenesis independent of tyrosine 705 phosphorylation. *Cancer Res* 2008; 68: 7736-7741.
26. Shariat SF, Lotan Y, Saboorian H, et al. Survivin expression is associated with features of biologically aggressive prostate carcinoma. *Cancer* 2004; 100: 751-757.

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